



EFFECT OF PHOSPHORUS FERTILIZER AND MYCORRHIZA ON PLANT HEIGHT, SEED WEIGHT PER PLANT AND GRAIN PER PANICLE IN WHEAT

Mohsen Noori^a, Alireza Sobhkhizi^a, Mohammad Adibian^a, Vahideh Mirikomak^a, Kabir Eyidozehl^{b*}

^aHigher Educational Complex of Saravan, Iran

^bYoung Researchers and Elite Club, Zahedan Branch, Islamic Azad University, Zahedan, Iran

Corresponding Author email: eyidozehl@gmail.com

ABSTRACT: Cereal crops occupy a prime position in providing food for human consumption. Arbuscular mycorrhizae are the most common type of mycorrhiza, where the fungi colonize the interior of the root and form specialized structures, known as arbuscules, for nutrient exchange with the host. In this study, research crops planted in 2011, and Khash mountain stage carried the gem industry. This study is a factorial experiment in a randomized complete block design with three replicates and all experiments were performed with different levels. In this experiment, a variety of wheat called clear that improved cultivars were used. According to the analysis of variance table mycorrhiza effect on Seed weight per plant, number of panicles per plant was significant.

Key words: panicles per plant, wheat, mycorrhiza

INTRODUCTION

Cereal crops occupy a prime position in providing food for human consumption and according to Graham and Welch [12] about 50% of the soil used for cereal production in the world contains low level of plant available zinc which reduces not only grain yield but also nutritional quality (low in micronutrients essential for good human health). Cereal grains are a major source of zinc intake for persons living in developing countries and zinc deficient cereal food is creating serious health problems. Seed zinc concentrations of wheat grown under zinc deficient conditions are very low [9]. Mycorrhizal fungi form a symbiotic relationship with plant roots. Arbuscular mycorrhizae are the most common type of mycorrhiza, where the fungi colonize the interior of the root and form specialized structures, known as arbuscules, for nutrient exchange with the host. Fungal hyphae extend from the root and explore the soil more efficiently than would fine plant roots. Arbuscular mycorrhizae can provide the plant with supplemental phosphorus (P), nitrogen (N), and micronutrients since the plant roots alone are not able to maximize the interception of nutrients [3]. One the most of important effects of mycorrhiza fungi is to increase crop yield, especially in soils with low fertility. Ortas [18] believes that use of the CGR mycorrhiza fungi and increase the allocation and transfer of materials between roots and stems of the leaves, so that more absorption of nutrients and transfer them to cause an increase in shoot dry weight. The yield increase may be due to the extension of roots, which penetrate through the mycelium of fungi in the soil and crops, access to a greater volume of soil [7] it also increases the tolerance of plant to drought stress [21]. Specifically in soybean, AM have been shown to improve the overall water status of the plant [19, 25] due to a reduced resistance to water transport [22], associated with an enhanced nutrient [22]. Numerous field studies have demonstrated the benefits of the mycorrhizal association to agricultural crops. Increased levels of root colonization and AMF hyphal density in soil at early growth stages can increase P uptake and yield in maize (*Zea mays* L.) when the soil is P deficient [5, 8, 11, 14]. Arbuscular mycorrhizae interact in the rhizosphere with other beneficial organisms and pathogens, often competing for the same colonization sites [13]. Mycorrhizae have been shown to protect plants against pathogens in numerous crops [4, 23, 26]. In soil with adequate plant available P, increased mycorrhizal fungus colonization has generally not translated into increased yields [10, 17, 24].

MATERIALS AND METHODS

In this study, research crops planted in 2011, and Khash mountain stage carried the gem industry. This study is a factorial experiment in a randomized complete block design with three replicates and all experiments were performed with different levels. In this experiment, a variety of wheat called clear that improved cultivars were used. Mycorrhiza arbuscular fungi (AM) in both the inoculated and non-inoculated with three levels of nitrogen and phosphorus fertilizer in three levels as other experimental treatments were used. Urea nitrogen is used by organizations of agricultural support services were provided. The farm has been in previous years under fallow land preparation including plowing, disk loader and fustigation is. The plowing by moldboard plow to a depth of 30 cm was used. The operation of the disc, the disc plow was perpendicular offset to a depth of 15 cm. To soil and plant nutrient land of the amount needed according to soil test results fustigation was done. To measure this trait after five plants were randomly selected and harvested from the middle two lines by removing the border took place clusters each of the plant to seed removed separately the for the plant out and counting were recorded. After data collection, by ANOVA statistical program SPSS, MASTATC took.

RESULTS AND DISCUSSION

Plant height

Analysis of variance was performed to test the interaction Mycorrhiza and P show a significant was not effect on wheat plant height (Table 1). Analysis of variance shows that the use of different levels of fertilizer (P) is causing a significant effect on plant height, so that the increase in chemical fertilizers (phosphorus) of plant height (Table 2). In this study, possibly because of the height, most of the plant, at the right time and the right amount of fertilizer for plants. It is well known that AM fungi are the causes of plant growth, mainly by nutrients uptake [6, 15]. Although AM fungi are generally considered to have a broad host ranges, some species are more effective with particular host plants such as corn, cotton and soybeans in increasing nutrients uptake, plant growth, water uptake and decrease negative effects of environment [20]. Phosphorus deficit is a most important restrictive factor in plant growth and recognition of mechanisms that increase plant phosphorus use efficiency is important [2].

Table 1. ANOVA analysis of the wheat affected by interactions of mycorrhiza in phosphorus

S.O.V	df	Plant height	Seed weight per plant	Grain per panicle	Number of panicles per plant
R	2	18.66	0.6	21.63	8.01
Mycorrhiza	2	1.50	42.04**	16.66	35.63**
P	2	34.56*	5.83	77.90*	7.18
P*M	2	18.2	16.54	87.5*	12.74
C.V	-	10.5	7.84	22.06	7.17
*, **, ns : significant at $p < 0.05$ and $p < 0.01$ and non-significant, respectively. P: Phosphorus, M: Mycorrhiza					

Seed weight per plant

According to the analysis of variance table mycorrhiza effect on grain yield per plant was significant (Table 1). According to the analysis of variance table Phosphorus effect on grain yield per plant was not significant (Table 1). The highest of the treated seed weight per 100 kilograms (20.46) and the lowest from the control treatment (15.6), respectively (Table 2). Grain yield and P accumulation by wheat were highest P rates [16]. The application of phosphorus fertilizer increased grain yield, at the growth stage [1].

Grain per panicle

According to the analysis of variance table mycorrhiza effect on grain per panicle was not significant (Table 1). According to the analysis of variance table Phosphorus effect on grain per panicle was significant (Table 1). The highest of the treated 100 kilograms (55.2) and the lowest from the control treatment (39.8), respectively (Table 2). The application of phosphorus fertilizer increased grain yield, at the growth stage [1]. Grain yield and P accumulation by wheat were highest P rates [16].

Table 2. Mean comparison of different characteristics influenced by mycorrhiza and phosphorus interactions.

Mean-square				
	Plant height	Seed weight per plant	Grain per panicle	Number of panicles per plant
inoculated				
0 kg	91.4	16.6	39.8b	7.6
50 kg	95.1	16.1	44.55b	8.7
100 kg	100.7	20.46	55.2a	11.4
Non-inoculated				
0 kg	94.4	15.6	40.3b	7
50 kg	96.11	17.2	43.4b	8
100 kg	97.7	18.4	51.3a	9.3

Any two means not sharing a common letter differ significantly from each other at 5% probability

Number of panicles per plant

According to the analysis of variance table mycorrhiza effect on number of panicles per plant was significant (Table 1). According to the analysis of variance table Phosphorus effect on number of panicles per plant was not significant (Table 1). The highest of the treated 100 kilograms (11.6) and the lowest from the control treatment (7.6), respectively (Table 2). In soil with adequate plant available P, increased mycorrhizal fungus colonization has generally not translated into increased yields [10, 17, 24].

REFERENCES

- [1] Alam MM, Hassanuzzaman M, Nahar K. 2009. Tiller dynamics of three irrigated rice varieties under varying phosphorus levels. *American – Eurasian Journal of Agronomy* 2 (2):89-94. (b).
- [2] Alinajati Sisie S, Mirshekari B. 2011. Effect of phosphorus fertilization and seed bio fertilization on harvest index and phosphorus use efficiency of wheat cultivars. *Journal of Food, Agricultural & Environment*. vol9 (2):388-397.
- [3] Allen, M.F., W. Swenson, J.I. Querejeta, L.M. Egerton-Warburton, and K.K. Treseder. 2003. Ecology of mycorrhizae: A conceptual framework for complex interactions among plants and fungi. *Annu. Rev. Phytopathol.* 41:271–303.
- [4] Azcon-Aguilar, C., and J.M. Barea. 1997. Arbuscular mycorrhizas and biological control of soilborne plant pathogens: An overview of the mechanisms involved. *Mycorrhiza* 6:457–464.
- [5] Boswell EP, Koide RT, Shumway DL, Addy HD 1998. Winter wheat cover cropping, VA mycorrhizal fungi and maize growth and yield. *Agr Ecosyst Environ* 67:55–65
- [6] Busse MD, Fiddler GO, Ratcliff AW, 2004. Ectomycorrhizal formation in herbicide treated soils of differing clay and organic matter content. *Water Air and Soil Pollut* 152: 23–34.
- [7] Cornejo P, Meier S, Borie G, Rillig MC, Borie F, *Sci. Total Environ*, 2008, 406:154-160.
- [8] Deguchi S, Shimazaki Y, Uozumi S, Tawaraya K, Kawamoto H, Tanaka O (2007) White clover living mulch increases the yield of silage corn via arbuscular mycorrhizal fungus colonization. *Plant Soil* 291:291–299
- [9] Erdal, I., Yilmaz, A., Taban, S., Ekar, S., Torun, B., Cakmak, I. 2002. Phytic acid and phosphorus concentrations in seeds of wheat cultivars grown with and without zinc fertilization. *Journal of Plant Nutrition* 25:113–127.
- [10] Galvez L, Douds DD, Wagoner P, Longnecker LR, Drinkwater LE, Janke RR 1995. An overwintering cover crop increases inoculum of VAM fungi in agricultural soil. *Am J Altern Agr* 10:152–156
- [11] Gavito ME, Miller MH 1998b. Early phosphorus nutrition, mycorrhizae development, dry matter partitioning and yield of maize. *Plant Soil* 199:177–186
- [12] Graham, A.W., McDonald, G.K. 2001. Effect of zinc on photosynthesis and yield of wheat under heat stress. *Proceedings of the 10th Australian Agronomy Conference 2001*, Australian Society of Agronomy. Hobart, Tasmania, Australia. Available on line at <http://www.regional.org.au/au/asa/2001/2/c/graham.htm>.

- [13] Harrier, L.A., and C.A. Watson. 2004. The potential role of arbuscular mycorrhizal (AM) fungi in the bioprotection of plants against soil-borne pathogens in organic and/or other sustainable farming systems. *Pest Manage. Sci.* 60:149–157.
- [14] Kabir Z, Koide RT 2000. The effect of dandelion or a cover crop on mycorrhiza inoculum potential, soil aggregation and yield of maize. *Agr Ecosyst Environ* 78:167–174
- [15] Karagiannidis N, Bletsos F, Stavropoulos N, 2002. Effect of verticillium wilt (*Verticillium dahliae* kleb.) and mycorrhiza (*Glomus mosseae*) on root colonization, growth and nutrient uptake in tomato and eggplant seedlings. *Sci Hort* 94: 145- 156.
- [16] Khan P, Aslam M, Momen MY, Imtiaz M, Shah JA, Depar N. 2010. Determining the Nutritional Requirements of Rice Genotype JAJAL 25/AEVOLED ANIA Tando Jam Pakistan. *Pakistan Journal Bot.* Vol 42 (24):3257-3263.
- [17] McGonigle TP, Miller MH 1999. Winter survival of extraradical hyphae and spores of arbuscular mycorrhizal fungi in the field. *Appl Soil Ecol* 12:41–50
- [18] Ortas I, *Sci. Plant. Anal*, 1996, 27: 2935-2946.
- [19] Porcel, R., and J.M. Ruiz-Lozano. 2004. Arbuscular mycorrhizal influence on leaf water potential solute accumulation, and oxidative stress in soybean plants subjected to drought stress. *J. Exp. Bot.* 55:1743–1750.
- [20] Powell JR, Campbell RG, Dunfield KE, Gulden RH, Hart MM, Levy-Booth, DJ, Klironomos J N, Pauls KP, Swanton CJ, Trevors JT, Antunes PM, 2009. Effect of glyphosate on the tripartite symbiosis formed by *Glomus intraradices*, *bradyrhizobium japonicum*, and genetically modified soybean. *Appl Soil Ecol* 41: 128-136.
- [21] Rillig MC, Mummey DL, Mycorrhizas and soil structure. *New Phytol*, 2006, 171: 41–53.
- [22] Safir, G.R., J.S. Boyer, and J.W. Gerdemann. 1972. Nutrient status and mycorrhizal enhancement of water transport in soybeans. *Plant Physiol.* 49:700–703.
- [23] Slezack, S., E. Dumas-Gaudot, M. Paynot, and S. Gianinazzi. 2000. Is a fully established arbuscular mycorrhizal symbiosis required for bioprotection of *Pisum sativum* roots against *Aphanomyces euteiches*? *Mol. Plant Microbe Interact.* 13:238–241.
- [24] Sorensen JN, Larsen J, Jakobsen I (2005) Mycorrhiza formation and nutrient concentration in leeks (*Allium porrum*) in relation to previous crop and cover crop management on high P soils. *Plant Soil* 273:101–114
- [25] Vejsadova, H., D. Siblikova, M. Gryndler, T. Simon, and I. Miksik. 1993. Influence of inoculation with *Bradyrhizobium japonicum* and *Glomus claroideum* on seed yield of soybeans under greenhouse and field conditions. *J. Plant Nutr.* 16:619–629.
- [26] Zhu, H.H., and Q. Yao. 2004. Localized and systemic increase of phenols in tomato roots induced by *Glomus versiforme* inhibits *Ralstonia solanacearum*. *J. Phytopathol.* 152:537–542.