



## RELATIONSHIP OF GRAIN FILLING WITH GRAIN YIELD OF MUNG-BEAN AFFECTED BY HYDRO-PRIMING DURATION AND WATER SUPPLY

Kazem Ghassemi-Golezani\*, Saeid Hassanpour-Bourkheili, Salar Farhanghi Abriz

Department of Plant Eco-Physiology, Faculty of Agriculture, University of Tabriz, Tabriz, Iran

E-mail: [golezani@gmail.com](mailto:golezani@gmail.com)

**ABSTRACT:** Grain filling is a crucial determinant of yield. Thus, a split-plot experiment based on RCB design with three replications was conducted in 2013, in order to evaluate the effects of water supply (I<sub>1</sub>, I<sub>2</sub>, I<sub>3</sub> and I<sub>4</sub>: irrigation after 70, 100, 130 and 160 mm evaporation from class A pan, respectively) and hydro-priming duration (P<sub>1</sub>, P<sub>2</sub> and P<sub>3</sub>: 0, 8 and 16 h, respectively) on grain filling of mung bean (*Vigna radiata* L). The highest grain filling rate was observed for P<sub>3</sub>, but there was no statistical difference between P<sub>1</sub> and P<sub>2</sub>. Grain filling duration and maximum grain weight decreased with decreasing water availability. Seeds hydro-primed for 16h had the highest maximum grain weight, but there was no significant difference between P<sub>2</sub> and P<sub>3</sub>. Water stress significantly reduced grain yield per unit area, although no significant difference between I<sub>1</sub> and I<sub>2</sub> treatments was observed. Grain yield per unit area for P<sub>3</sub> plants was higher than that of P<sub>1</sub> and P<sub>2</sub> plants. Grain filling duration and maximum grain weight had the highly positive correlations with grain yield per unit area. Hydro-priming for 16 h had the highest beneficial effects on field performance of mung-bean seeds.

**Key words:** Grain filling, Grain yield, Mung-bean, Water stress

### INTRODUCTION

Mung-bean (*Vigna radiata* L.) is one of the most nutritious grain legumes used in different parts of the world [1]. Although this plant is a relatively drought tolerant crop and performs well under conditions of low soil moisture [2], severe water stress can result in yield loss [3]. Drought stress is the most common form of abiotic stress and plants are likely to encounter periods of water shortage in their life cycle [4]. Water is essential to plant growth because it provides the medium within which most cellular functions take place [5]. Water stress causes membrane damage, and stimulates molecular signal transduction and hormone activation [6], leading to a reduction in plant growth and productivity [7, 8, 9, 10]. The degree of yield reduction is determined by the timing, severity and duration of the water deficit. Water stress during vegetative stages has the greatest impact on plant height and biomass [11], but during the reproductive growth it is considered to have the most adverse effect on crop productivity [12]. Grain filling is a crucial determinant of yield and is characterized by duration and rate of filling [13]. Water stress occurring during grain development curtails the kernel sink potential by reducing the number of endosperm cells and amyloplasts formed [14], thus reducing grain weight as a result of a reduction in the capacity of the endosperm to accumulate starch [15]. Some of the deleterious effects of environmental stresses such as water stress on crop performance may be overcome by seed priming [16, 9]. Seed priming is the soaking of seeds in a solution of any priming agent followed by drying of seeds that initiates germination related processes without radicle emergence [17]. Various seed priming techniques have been developed, including hydro-priming (soaking in water), halo-priming (soaking in inorganic salt solutions), osmo-priming (soaking in solutions of different organic osmotica), thermo-priming (treatment of seeds with low or high temperatures), matrix priming (treatment of seed with solid matrices) and bio-priming (hydration using biological compounds) [18]. Seed invigoration by priming treatments has been associated with various biochemical, cellular and molecular events including synthesis of RNA and proteins [19, 20]. Priming also restores activities of enzymes involved in the cell detoxifying mechanisms [21]. Earlier works showed that the success of seed priming is influenced by the complex interaction of factors including plant species, water potentiality of priming agent, duration of priming, temperature, seed vigor and storage conditions of the primed seeds [22]. The beneficial effects of seed priming have been demonstrated for many field crops such as pinto [23], rapeseed [24], borage [25], chickpea [26] and lentil [27]. This research was conducted to evaluate the effects of hydro-priming duration on grain filling of mung-bean under different irrigation treatments.

## MATERIALS AND METHODS

The experiment was conducted in 2013 at the Research Farm of the Faculty of Agriculture, University of Tabriz, Iran (Latitude 38°05' N, Longitude 46°17' E, Altitude 1360 m above sea level). Seeds of mung-bean (*vigna radiata* L.) were divided into three sub-samples, one of which was kept as control (non-primed, P<sub>1</sub>) and two other samples were soaked in distilled water at 20°C for 8 (P<sub>2</sub>) and 16 (P<sub>3</sub>) hours and then dried back to initial moisture content at room temperature of 20-22°C. All the seeds were treated with benomyl at a rate of 2 g kg<sup>-1</sup> before sowing. Seeds were hand sown in about 4 cm depth with a density of 80 seeds per m<sup>2</sup> on 7 May 2013. Each plot consisted of 6 rows with 3.5 m length, spaced 25 cm apart. The experiment was arranged as split-plot, based on RCB design with three replications. All plots were irrigated immediately after sowing and subsequent irrigations were carried out after 70 (I<sub>1</sub>), 100 (I<sub>2</sub>), 130 (I<sub>3</sub>) and 160 (I<sub>4</sub>) mm evaporation from class A pan. Weeds were frequently controlled by hand during crop growth and development.

Grains were harvested in five day intervals at eight stages. The grains of each harvest were separately dried in an oven at 80°C for 48 hours, and then weighed. Grain filling rate was calculated by using the following equation:

$$\text{Grain filling rate (mg d}^{-1}\text{)} = \text{Maximum grain weight (mg)} / \text{Grain filling duration (day)}$$

At maturity, plants in 1 m<sup>2</sup> of the middle part of each plot were harvested and grain yield per unit area was determined. Analysis of variance of the data appropriate to the experimental design and comparison of means at  $p \leq 0.05$  were carried out, using MSTATC and SPSS softwares.

## RESULTS AND DISCUSSION

Analysis of variance indicated that hydro-priming significantly influenced grain filling rate, but there was no significant difference among irrigation treatments. However, grain filling duration was significantly affected by irrigation treatments and hydro-priming durations had no significant effect on this trait. Maximum grain weight and grain yield were significantly affected by both hydro-priming and irrigation (Table 1).

The highest grain filling rate was observed in P<sub>3</sub>, However, there was no statistical difference between P<sub>1</sub> and P<sub>2</sub> (Table 2). This may be due to enhanced endosperm cells division and cytokinin activities resulting in improved grain filling rate. Pre-sowing treatment reduces accumulation of abscisic acid (ABA) and accelerates the grain filling rate [28].

Severe water stress levels significantly shortened the duration of grain filling. The highest grain growth period was observed under I<sub>1</sub>, but it was statistically similar with I<sub>2</sub> (Table 2). Environmental stresses such as water shortages, especially during grain filling, cause reductions in photosynthesis and remobilization of stored materials and hence, grain filling duration [29]. Water stress generally accelerates leaf senescence and shortens grain filling duration [30]. Similar results were reported for faba-bean [10] and chickpea [31]. Maximum grain weight decreased with increasing drought stress. The highest and the lowest maximum grain weights were obtained under I<sub>1</sub> and I<sub>4</sub>, respectively (Table 2). Maximum grain weight of mung-bean seeds was mainly influenced by grain filling duration rather than by grain filling rate, which is also reflected in the highest positive correlation of grain filling duration with maximum grain weight (Table 3). These results are strongly supported by the work of Ghassemi-Golezani *et al.* [10] on faba-bean. Seeds hydro-primed for 16h had the highest maximum grain weight, but there was no significant difference between P<sub>2</sub> and P<sub>3</sub> (Table 2). This may be due to higher sink activity in plants derived from primed seeds, which in turn is caused by more activity of enzymes involved in sucrose [32]. Bakht *et al.* [33] also reported that primed seeds produce plants with larger grains.

**Table 1. Analysis of variance of the effects of hydro-priming duration on grain filling, maximum grain weight, and grain yield of mung-bean**

Source of Variation	df	MS			
		Grain filling rate	Grain filling duration	Maximum grain weight	Grain yield
Replication	2	0.006 <sup>ns</sup>	2.282 <sup>ns</sup>	0.759 <sup>ns</sup>	198.347 <sup>ns</sup>
Irrigation (I)	3	0.004 <sup>ns</sup>	95.120 <sup>**</sup>	248.174 <sup>**</sup>	8072.477 <sup>**</sup>
Error a	6	0.002	0.923	0.681	134.201
Hydro-priming (HP)	2	0.01 <sup>*</sup>	0.949 <sup>ns</sup>	6.783 <sup>**</sup>	3153.889 <sup>*</sup>
I × HP	6	0.001 <sup>ns</sup>	0.769 <sup>ns</sup>	0.796 <sup>ns</sup>	72.836 <sup>ns</sup>
Error b	16	0.002	0.764	0.781	516.084
CV		2.79%	2.16%	1.5%	15.8%

ns,\* and \*\* : No significant and significant at  $p \leq 0.05$  and  $p \leq 0.01$ , respectively

**Table 2. Means of mung-bean field traits influenced by irrigation and hydro-priming duration**

Treatments	Grain filling rate (mg d <sup>-1</sup> )	Grain filling duration (day)	Maximum grain weight (g)	Grain yield (g m <sup>-2</sup> )
<b>Irrigation</b>				
I <sub>1</sub>	1.473 a	43.386 a	63.863 a	174.7 a
I <sub>2</sub>	1.461 a	42.846 a	62.561 b	162.3 a
I <sub>3</sub>	1.477 a	38.899 b	57.436 c	128.2 b
I <sub>4</sub>	1.433 a	36.584 c	52.370 d	109.9 c
<b>Hydro-priming</b>				
P <sub>1</sub>	1.437 b	40.607 a	58.345 b	126.9 b
P <sub>2</sub>	1.454 b	40.574 a	58.984 b	145.2 ab
P <sub>3</sub>	1.492 a	40.105 a	59.843 a	159.2 a

Different letters at each column for each treatment indicate significant difference at  $p \leq 0.05$ .

I<sub>1</sub>, I<sub>2</sub>, I<sub>3</sub>, I<sub>4</sub>: irrigation after 70, 100, 130 and 160 mm evaporation from class A pan, respectively

P<sub>1</sub>, P<sub>2</sub> and P<sub>3</sub>: non-primed and hydro-primed seeds for 8 and 16 h, respectively

**Table 3. Correlation coefficients of traits**

Trait	1	2	3	4
1- Grain filling rate	1			
2- Grain filling duration	0.233	1		
3- Maximum grain weight	0.486	0.963**	1	
4- Grain yield	0.567	0.848**	0.920**	1

\*\* : Statistically significant at  $p \leq 0.01$

Water stress significantly reduced grain yield per unit area, although no significant difference between I<sub>1</sub> and I<sub>2</sub> treatments was observed (Table 2). Water limitation during flowering stage leads to flower abortion, poor pod set and formation of infertile pods, which can potentially reduce grain yield per unit area [34]. Grain yield per unit area for P<sub>3</sub> plants was higher than that of P<sub>1</sub> and P<sub>2</sub> plants, but P<sub>3</sub> plants had no significant difference with P<sub>2</sub> plants (Table 2). Harris *et al.* [35] found that hydro-priming resulting in faster development, earlier flowering and maturity and higher yields of upland rice, maize and chickpea. The resultant effects of priming depend on duration of seed soaking, beyond which it could be damaging to the seed or seedling [36]. Optimal times of hydro-priming were 7 h for pinto bean [23], 12 h for chickpea [26], and 8 h for lentil [27]. In our research, the best duration for hydro-priming of mung-bean seeds was 16 h. Correlation of grain filling duration with maximum grain weight and correlations of grain filling duration and maximum grain weight with grain yield were positive and significant. However, grain filling rate had no significant correlation with other traits. Thus, grain filling duration and maximum grain weight have the major roles in determining grain yield of mung-bean.

## REFERENCES

- [1] Duke, J.A. 1981. Handbook of legumes of world economic importance. Plenum Press, New York. pp: 345.
- [2] Lalinia, A.A., Majnon Hoseini, N., Galostian, M., Esmailzadeh Bahabadi, S., Marefatzadeh Khameneh, M. 2012. Echophysiological impact of water stress on growth and development of mungbean. Intl J Agron Plant Prod, 3, pp. 599-607.
- [3] Srivastava, R.L., Sahai, R.N.J., Axena, K.S., Singh, I.P. 1976. Path analysis of yield component in soybean. Indian J Agr Res, 10, pp. 171-173.
- [4] Cruz de Carvalho, M.H. 2008. Drought stress and reactive oxygen species: Production, scavenging and signaling. Plant Signal Behav, 3, pp. 156-165.
- [5] Condon, A.G., Richards, R.A., Rebetzke, G.J., Farquhar, G.D. 2004. Improving intrinsic water use efficiency and crop yield. Crop Sci, 42, pp. 122-131.
- [6] Knight, H., Knight, M.R. 2001. Abiotic stress signaling pathways: specificity and cross-talk. Trends Plant Sci, 6, pp. 262-267.
- [7] Saranga, Y., Menz, M., Jiang, C.X., Wright, R.J., Yakir, D., Paterson, A.H. 2001. Genomic dissection of genotype x environment interactions affecting genetic analysis of root response to drought stress and abscisic acid. Plant Physiol, 142, pp. 1065-1074.

- [8] Showler, A.T., Moran, P.J. 2003. Effects of drought stressed cotton, *Gossypium hirsutum L.*, on beet armyworm, *Spodoptera exigua* (Hubner), oviposition, and larval feeding preferences and growth. *J Chem Ecol*, 29, pp. 1997-2011.
- [9] Ghassemi-Golezani, K., Sheikhzadeh-Mosaddegh, P., Valizadeh, M. 2008. Effects of hydro-priming duration and limited irrigation on field performance of chickpea. *Res J Seed Sci*, 1, pp. 34-40.
- [10] Ghassemi-Golezani, K., Ghanehpour, S., Dabbagh Mohammadi-Nasab, A. 2009. Effects of water limitation on growth and grain filling of faba bean cultivars. *J Food Agric Environ*, 7, pp. 442-447.
- [11] Ghassemi-Golezani, K., Andalibi, B., Zehtab-Salmasi, S., Saba, J. 2008. Effect of water stress during vegetative and reproductive stages on seed yield and essential oil content of dill (*Anethum graveolens L.*). *J Food Agri Environ*, 6, pp. 282-284.
- [12] Costa-Franca, M.G., Thi, A.T., Pimental, C., Pereyra, R.O., Zuily-Fodil, Y., Laffray, D. 2000. Differences in growth and water relations among *Phaseolus vulgaris L.* cultivars in response to induced drought stress. *Environ Exp Bot*, 43, pp. 227-237.
- [13] Yang, W., Peng, S., Dionisio-Sese, M.L., Laza, R.C., Visperas, R.M. 2008. Grain filling duration, a crucial determinant of genotypic variation of grain yield in field-grown tropical irrigated rice. *Field Crops Res*, 105, pp. 221-227.
- [14] Saini, H.S., Westgate, M.E. 2000. Reproductive development in grain crops during drought. *Adv Agron*, 68, pp. 59-95.
- [15] Nicolas, M.E., Lambers, H., Simpson, R.J., Dalling, M.J. 1985. Effect of drought on metabolism and partitioning of carbon in two wheat varieties differing in drought-tolerance. *Ann Bot*, 55, pp. 727-747.
- [16] Demir Kaya, M., Okcu, G., Atak, M., Cikili, Y., Kolsarici, O. 2006. Seed treatments to overcome salt and drought stress during germination in sunflower (*Helianthus annuus L.*). *Eur J Agron*, 24, pp. 291-295.
- [17] McDonald, M.B., 1999. Seed deterioration: physiology, repair and assessment. *Seed Sci Technol*, 27, pp. 177-237.
- [18] Ashraf, M., Foolad, M.R. 2005. Pre-sowing seed treatment: A shotgun approach to improve germination, plant growth and crop yield under saline and non-saline conditions. *Adv Agron*, 88, pp. 223-271.
- [19] Dell'Aquila, A., Bewley, J.D. 1989. Protein synthesis in the axes of polyethylene glycol treated pea seeds and during subsequent germination. *J Exp Bot*, 40, pp. 1001-1007.
- [20] Davison, P.A., Bray, C.M. 1991. Protein synthesis during osmo-priming of leek (*Allium Porrum L.*) seeds. *Seed Sci Res*, 1, pp. 29-35.
- [21] Bailly, C., Benamar, A., Corbineau, F., Come, D. 1997. Changes in superoxide dismutase, catalase and glutathione reductase activities in sunflower seeds during accelerated ageing and subsequent priming. In: Ellis (eds). *Basic and applied aspects of seed biology*. Dordrecht: Kluwer Academic Publishers, pp: 665-672.
- [22] Parera, C.A., Cantliffe, D.J. 1994. Pre-sowing seed priming. *Hort Rev*, 16, pp. 109-141.
- [23] Ghassemi-Golezani, K., Chadordooz-Jeddi, A., Nasrullahzadeh, S., Moghaddam, M. 2010. Influence of hydro-priming duration on field performance of pinto bean (*Phaseolus vulgaris L.*) cultivars. *Afr J Agric Res*, 5, pp. 893-897.
- [24] Ghassemi-Golezani, K., Jabbarpour, S., Zehtab-Salmasi, S. 2013. Influence of seed priming on ground cover and grain yield of spring and winter rapeseed cultivars. *Int J Biosci*, 3, pp. 54-61.
- [25] Ghassemi-Golezani, K., Dastborhan, S., Zehtab-Salmasi, S. 2013. Seed Priming and Field Performance of Borage (*Borago officinalis L.*) under Different Irrigation Treatments. *Intl J Agron Plant Prod*, 4, pp. 82-87.
- [26] Ghassemi-Golezani, K., Hosseinzadeh-Mahootchy, A., Zehtab-Salmasi, S., Turchi, M. 2013. Influence of seed invigoration and water supply on morpho-physiological traits of chickpea. *Intl J Agron Plant Prod*, 4, pp. 782-786.
- [27] Ghassemi-Golezani, K., Jabbarpour-Bonyadi, Z., Shafagh-Kolvanagh, J., Nikpour-Rashidabad, N. 2013. Effects of Water Stress and hydro-priming duration on field performance of lentil. *Intl J Farm & Alli Sci*, 2013, 2, pp. 922-925.
- [28] Sakhabutdinova, A.R., Fatkutdinova, D.R., Bezrukova, N.V., Shakirova, F.M. 2003. Salicylic acid prevents the damaging action of stress factors on wheat plants. *Bulg J Plant Physiol*, 21, pp. 314-319.
- [29] Sadeghipour, O. 2008. Effect of withholding irrigation at different growth stages on yield and yield components of mung bean (*Vigna radiata L. Wilczek*) varieties. *Am-Euras J Agric & Environ Sci*, 4, pp. 590-594.
- [30] Davis, S., Turner, N.C., Siddique, K.H.M., Lepore, L., Plummer, J. 1999. Seed growth of Desi and Kabuli chickpea (*Cicer arietinum L.*) in a short season Mediterranean-type environment. *Aust J Exp Agric*, 39, pp. 181-188.
- [31] Mustafavi, S.H., Ghassemi-Golezani, K., Shafagh-Kalvanagh, K., Movludi, A. 2013. Effect of irrigation disruption at reproductive stages on grain filling of chickpea cultivars. *Intl J Agron Plant Prod*, 4, pp. 863-868.

- [32] Kaur, S., Gupta, A.K., Kaur, N. 2005. Seed Priming Increases Crop Yield Possibly by Modulating Enzymes of Sucrose Metabolism in Chickpea. *J Agron Crop Sci*, 191, pp. 81–87.
- [33] Bakht, J., Shah, R., Shafi, M., Amankhan, M. 2010. Effect of various priming sources on yield and yield components of maize cultivars *Pak J Bot*, 42, pp. 4123-4131.
- [34] Fang, X., Turner, N.C., Yan, G., Li, F., Siddique, K.H.M. 2009. Flower numbers, pod production, pollen viability, and pistil function are reduced and flower and pod abortion increased in chickpea (*Cicer arietinum L.*) under terminal drought. *J Exp Bot*, 61, pp. 335–345.
- [35] Harris, D., Joshi, A., Khan, P.A., Gothkar, P., Sodhi, P.S. 1999. On-farm seed priming in semi-arid agriculture: Development and evaluation in maize, rice and chickpea in India using participatory methods. *Exp Agric*, 35, pp. 15-29.
- [36] Kumar, A., Gangwar, J.S., Prasad, S.C., Harris, D. 2002. On-farm seed priming increases yield of direct-sown finger millet in India. *International Sorghum & Millets Newsletter*, 43, pp. 90-92.