

LIGATION OF E2 GENE WITH REPLICASE BASED EUKARYOTIC pSinCMV VECTOR AND ITS TRANSFORMATION STUDY

Nitin Sharma*, Purshotam Kaushik¹, Anant Rai²

*Corresponding author: email- nitinsharma.com@gmail.com, Mb no,-09897857585

¹Dept of Microbiology, Gurukul Kangri Vishwavidyalaya-Haridwar

²Institute of Biotechnology & IT, 197, Mudia Ahamadnagar, Pilibhit Road, Bareilly (UP).

ABSTRACT: The E₂ gene of classical swine fever virus (CSFV) is already cloned in pVAX1 vector. In the present study we have released the E₂ gene insert from the pVAX1.CSFV.E₂ using plasmid isolation, RE digestion and ligated with pSinCMV vector. Prepared the competent cell to transformation of Ligated product in *E.coli* DH5 α . Make the L.B. Agar plate and the large number of colonies (approximately 25) were picked from the overnight grown transformants. The individual colonies were inoculated in fresh ampicillin (50 μ g/ml) containing LB broth and allowed to grow for large scale production for further experiments. This can be used for expression and immunological studies.

Key words: Classical Swine Fever Virus, Transformation, E₂ gene, Restriction enzyme.

INTRODUCTION

Classical swine fever (CSF), also known as hog cholera, is a highly contagious, multisystemic and hemorrhagic viral disease, included in the list of diseases notifiable to the OIE (www.oie.int), distributed almost world-wide and is considered the most economically important but vaccinepreventable disease of swine in areas of intensive pig farming. Natural host of classical swine fever virus are members of the family Suidae, which include domestic pigs and wild boars (Depner et al., 1995; Laddomada, 2000). A considerable problem is the survival CSF virus in wild pig population, which is considered to be the potential source of the infection.

Genomic RNA possess single ORF. Initially a polyprotein is formed which is further cleaved cotranslationally and post-translationally by the host and viral genome encoded proteases to yield four structural (C protein, Erns, E1 and E2) and seven non structural proteins. Erns, E1 and E2 are the viral envelope glycoprotein. After infection with CSFV, antibodies are raised against the structural glycoproteins E2 and Erns and non structural protein NS3 (Paton et al., 1991; Terpstra, 1988). Antibodies against Erns and NS3 have only low neutralizing capacity or none at all (Rumenapf et al., 1991; Konig et al., 1995).

E2 glycoprotein contains most of the known humoral and cell mediated protective determinants of CSFV (Ceppi et al., 2005), highly immunogenic against which most of the neutralizing antibodies are induced and the only one capable of conferring protection against CSFV challenge (Rumenapf et al., 1991; Hulst et al., 1993).

MATERIALS AND METHODS

Vector: The replicase based pSinCMV vector (Nagarajan, 2005), was used.

CSFV E₂ gene

The E₂ gene already cloned in pVAX1 vector was used as the candidate gene, the sequence accession no. EU857642.

Host Bacterial strains

Escherichia coli (E.coli) DH5 α (Proteges, Madison, USA) host strain was used for transformation experiments.

Preparation for E₂ gene

pVAX1.CSFV.E₂ recombinant plasmid containing E₂ gene insert was revived in LB broth and plasmid DNA was isolated (Sambrook and Russell 2001). Restriction endonuclease digestion of recombinant plasmid pVAX1.csfv.E₂ with PmeI enzyme was done to release the E₂ gene insert. This enzyme create blunt end. The size of E₂ gene ORF is 1.125 Kb.

pSinCMV vector preparation and ligation

StuI was use to create blunt end using 50 μ l reaction mix. A 10 μ l reaction mixture was for blunt end ligation containing T4 DNA ligase (Fermentas) 1 μ l, pSinCMV Vector 2 μ l, CSFV E₂ gene 5 μ l, ligation buffer (10X) 1 μ l, nuclease free water 1 μ l. The reaction mixture was incubated overnight at 14°C. The linearised plasmid was checked and quantitated on 1% agarose gel electrophoresis. The size of pSinCMV vector is 10.779 Kb.

Preparation of competent cells and Transformation

Took the 500 μ l and equal volume of ice-cold 2xTSS is added in tube and the cell suspension mixed gently. Then 2 μ l Ligated DNA and mixed the 100 μ l cell suspension (competent cell), mixture incubate for 5-6 min at 4°C, add the 0.9 ml of SOC, incubate 37°C with shaking for 1 hr to allow expression of the antibiotic-resistance gene. Transformants are selected by standard methods (Chung et al., 1989).

Screening of recombinant clones

A large number of colonies (approximately 25) were picked from the overnight grown transformants. The individual colonies were inoculated in fresh ampicillin (50 μ g/ml) containing LB broth and allowed to grow for 18 to 24 hours. This culture can be use for immunological studies.

RESULTS**Subcloning of csfv E₂ gene in pSinCMV.csfv. E₂**

The csfv E₂ gene cloned in pVAX1 vector was released by digesting with PmeI enzyme then makes the 2 fragments one is vector's size 2979 bp and second is 1119 bp of E₂ gene (**Fig. 1**). This PmeI enzyme create blunt end. The pSinCMV vector was linearized by digesting with StuI enzyme for used to create blunt end size 10779 bp (**Fig. 2**), gel eluted, purified by phenol chloroform precipitation method. Blunt end ligation was done and ligated vector was transformed in *E.Coli* (DH5 α) cells. Transformed product was spread in L.B. agar plate, after incubated over night at 37°C blue-white colony seen (**Fig. 3**).

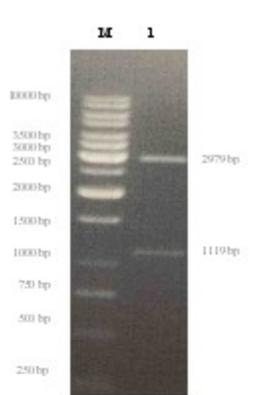


Fig 1. Restriction digestion of pVAC1. Lane M- Marker 1 Kb Ladder Lane 1- Digestion with PvuII releasing a product of 2979 bp & 1119 bp

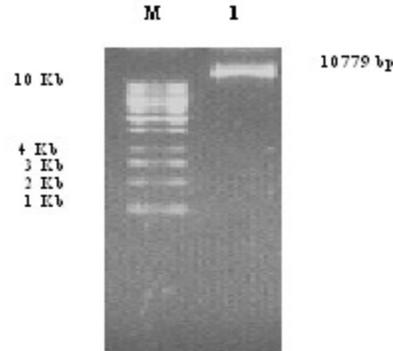


Fig 2. Preparation of pSinCMV vector Line M- 1 Kb DNA Marker Line 1- pSinCMV vector digestion with StuI enzyme

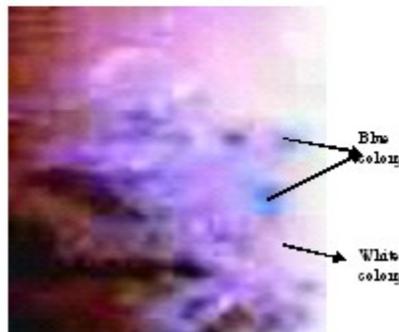


Fig 3. Transformation show Blue & White colony in plate

DISCUSSION

E₂ gene has become the main candidate for the development of DNA vaccine against CSFV. We have tried to transform the E₂ gene in replicase based eukaryotic vector namely pSinCMV.

pSinCMV is an alpha virus (Sindbis virus) based plasmid vector which serves as negative strand once it enters into the host cell. There after entering into the nucleus it is transcribed by host RNA polymerase enzymes from CMV promoter into a full length RNA transcript. This full length transcript then acts as positive sense alpha virus which in turn is translated in cytoplasm to form replicase protein. This protein serves as RNA dependent RNA polymerase enzyme and forms negative sense RNA from positive sense transcript. From this negative sense strand full length as well as smaller fragments from subgenomic promoters are transcribed which in-turn is translated into proteins.

Since the cloned insert is downstream to the subgenomic promoter, the translated proteins represents our target proteins. The subgenomic promoter of alpha virus is very strong so that it makes large number of target mRNA from the sequence downstream to it. Replicase based vectors are superior over other conventional vectors in terms of its lower requirement of dose of immunization (Xiong et al., 1989; Hariharan et al., 1998, Berglund et al., 1998, Leitner et al., 2000).

In the transformation blue-white colony showed the E₂ gene is transformed in pSinCMV vector on the plate. Further we can use of pSinCMV.csfv.E₂ for large scale production in L.B. broth and after confirmation by PCR and sequencing can be use in expression and immunological study.

Acknowledgement

I am extremely grateful to my Parent for motivate me in this work.

REFERENCES

- Berglund P., Smerdou C., Fleeton M.N., Tubulekas I., Liljestrom P. (1998) Enhancing immune responses using suicidal DNA vaccines. *Nat. Biotechnol.* 16:562–565.
- Cepi M., De Bruin M.G., Seuberlich T., Balmelli C., Pascolo S., Ruggli N., Wienhold D., Tratschin J.D., McCullough K.C., Summerfield A. (2005). Identification of classical swine fever virus protein E2 as a target for cytotoxic T cells by using mRNA transfected antigenpresenting cells. *Journal of General Virology*, 86: 2525–2534.
- Depner K.R., Muller A., Gruber A., Rodriguez A., Bickhardt K., Liess B.(1995). Classical swine fever in wild boar (*Sus scrofa*)-experimental infections and viral persistence. *Dtsch. Tierarztl. Wochenschr.* 102:381-384.
- Hariharan M.J., Driver D.A., Townsend K., Brumm D., Polo J.M., Belli B.A., Catton D.J., Hsu D., Mittelstaedt D., McCormack J.E., Karavodin J., Dubensky T.W., Chang S.M. Jr., Banks T.A. (1998). DNA immunization against herpes simplex virus: enhanced efficacy using a Sindbis virusbased vector. *J. Virol.*, 72:950–958.
- Hulst M.M., Westra D.F., Wensvoort G., Moormann R.J.M. (1993). Glycoprotein E1 of hog cholera virus expressed in insect cells protects swine from hog cholera. *Journal of Virology*, 67: 5435-5442.
- Konig M., Lengsfeld T., Pauly T., Stark R., Thiel. H.J.(1995). Classical swine fever virus: independent induction of protective immunity by two structural glycoproteins. *Journal of Virology*, 69: 6479–6486
- Laddomada A. (2000). Incidence and control of classical swine fever in European wild boar. *Veterinary Microbiology*, 73: 121-130.
- Leitner W.W., Ying H., Driver D.A., Dubensky T.W., Restifo, N.P. (2000). Enhancement of tumor specific immune response with plasmid DNA replicon vectors. *Cancer Res.*, 60: 51-55.
- Paton D. J., Ibata G., Edwards S., Wensvoort G. (1991). An ELISA detecting antibodies to conserved pestivirus epitopes. *Journal of Virological Methods*, 31: 315-324.
- Rumenapf T., Stark R., Meyers G., Thiel H.J.(1991). Structural proteins of hog cholera virus expressed by vaccinia virus: further characterization and induction of protective immunity. *Journal of Virology*, 65: 589–597.
- Sambrook J., Russel D.W. 2001. *Molecular cloning: A laboratory manual*. Cold Spring Harbor laboratory press, Cold spring harbor, New York.
- Terpstra C., Wensvoort G.(1988). The protective value of vaccine induced neutralizing antibody titres in swine fever. *Veterinary Microbiology*, 16: 123-128.
- Xiong C., Levis R., Shen P., Schlesinger S., Rice C.M., Huang H.V. (1989). Sindbis virus: an efficient, broad host range vector for gene expression in animal cells. *Science*, 243:1188–1191.